adjacent mercurated calixarenes at the air-water interface. The fact that malonic and not oxalic acid is capable of increasing the cohesiveness of the calix[6]arene monolayer strongly suggests that bridging is, in fact, responsible for this stabilization.¹

Direct evidence for the porous character of compressed monolayers of 2b (before and after treatment with malonate) has been obtained by measurement of water evaporation through the film.17 Placement of a Ni/Cr wire basket, containing 200 mg of dried (170 °C) 6-12 mesh silica desiccant, 5 mm above a monolayer of 2b for 1 h (20 dyn/cm; 20 µM sodium trifluoroacetate subphase, maintained at 25 °C), resulted in water evaporation which was $92.4 \pm 5.6\%$ of that found with use of a clean surface; with a 10 μ M malonic acid subphase, the percentage was 88.4 ± 3.6%.^{18,20} Similar experiments that were carried out by using a monolayer of 1-hexadecanol (20 dyn/cm; 20 µM sodium trifluoroacetate subphase) showed a water evaporation of $64.0 \pm 3.5\%$. Thus, monolayers of 2b, compressed to a surface pressure of 20 dyn/cm, maintain a pore structure which offers relatively little resistance toward the permeation of water.21

Studies that are now in progress are aimed at fabricating composite membranes based on perforated monolayers derived from 2a-d, defining their permeability characteristics, and synthesizing related "porous surfactants" for use in the construction of other perforated monolayers.

(18) The percentage of water evaporation relative to bare surface is defined as $\left[(A - C)/(B - C) \right] \times 100$, where A and B are the percent weight gains in the presence and absence of the monolayer, respectively, and C is the percent weight gain in the laboratory ambient.¹⁹ (19) Archer, R. J.; LaMer, V. K. J. Phys. Chem. **1955**, 59, 200.

(20) All evaporation data are reported as averages of at least four inde-pendent experiments, carried out with a MG Lauda film balance.

(21) CPK models predict a maximum pore diameter of 6.4 Å, when each of the aromatic rings are approximately perpendicular to the air-water interface. Rotation about the bridging methylenes can further reduce the effective pore size.

The Preparation and Crystal Structures of New Platinum/Phosphonate Complexes

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Certain phosphonates [e.g., phosphonoformic acid (PFA), phosphonoacetic acid (PAA), some derivatives of methylene diphosphonate (MDP), and others] are known to inhibit Herpes, influenza, and other viruses.¹⁻³ Their anti-viral activity is associated with their inhibition of viral polymerases and transcriptases,4-6 dication-dependent enzymes which catalyze the



Figure 1. Structure of the [cis-Pt(NH₃)₂(PFA)]⁻ anion. Average distances around the periphery of the Pt atom are normal: Pt-N = 2.09(2), Pt-O(C) = 2.08 (1), C-O = 1.26 (2), C-P = 1.85 (2), P-O = 1.56(2), and Pt-O(P) = 2.05 (2) Å. $Na^+[cis-Pt(NH_3)_2Pt(PFA)^-3H_2O$ (2), and the tricline space group P1, with a = 6.366 (8) Å, b = 13.00 (2) Å, c = 6.933 (6) Å, $\alpha = 93.30$ (9)°, $\beta = 114.22$ (8)°, $\gamma = 93.96$ (10)°, $\rho_c = 2.703$ g cm⁻³ for Z = 2. The structure was solved by standard heavy atom techniques and refined a final agreement factor of R = 3.4%, R(w) = 5.8% for 833 reflections. A listing of final atomic coordinates is available.15



Figure 2. Structure of Na⁺[Pt(*trans-l*-dach)(PFA)]⁻. The compound crystallizes in the cubic space group P432, with a = 21.631 (8) Å, V =10120 (7) Å³, Z = 24. The structure was solved by using direct methods to locate the platinum atom and refined to present agreement factors of R = 9.9% and R(w) = 7.7% for 842 reflections.

formtion of a complementary strand of polynucleotide from a single-stranded parent polynucleotide. $^{6-9}$ The molecular basis of phosphonate action has not been unambiguously established, but it is strongly suspected tht the phosphonates bind either to an endogenous metal of the enzyme (e.g., Zn²⁺) or the exogenous coenzymes (e.g., Mg²⁺ or Mn²⁺) required for enzymatic activity. Thus, the paucity of information about the ligand properties of these compounds, and the structures of their complexes, is somewhat surprising. Only a few structures of MDP¹⁰ and py-

(5) Vrang, L.; Oberg, B. Antimicrob. Agents Chemother. 1986, 29, 867.

⁽¹⁶⁾ Monolayers of 2b have also been stabilized using dithiothreitol (10 (16) Monologies of 20 marcals occurs are been structure in the dimension of the μ M). In this case, surface pressures fell from 25.4 to 24.7 dyn/cm after 71 min and remained unchanged after 131 min. Transfer of these monolayers to glass slides, however, gave irreproducible transfer ratios that ranged between 0 and 0.9.

⁽¹⁷⁾ Adamson, A. W. Physical Chemistry of Surfaces, 4th ed.; John Wiley: New York, 1982; p 143.

^{(1) (}a) Herrin, T. R.; Fairgrieve, J. S.; Bower, R. R.; Shipkowitz, N. L.; Mao, J. C.-H. J. Med. Chem. 1977, 20, 5. (b) Mao, J. C.-H.; Otis, E.; von Esch; Herrin, T. R.; Fairgrieve, J. S.; Shipkowitz, N. L.; Duff, R. G. Antim-icrob. Agents Chemother. 1985, 27, 197.

^{(2) (}a) Oberg, B. Pharmac. Ther. 1983, 9, 387. (b) Sarin, P. S.; Taguchi, Y.; Sun, D.; Thornton, A.; Gallo, R. C.; Oberg, G. Biochem. Pharmacol. 1985, 34, 4075

⁽³⁾ Hutchinson, D. W. Anti-viral Res. 1985, 5, 193.
(4) (a) Chang, Y. C.; Grill, S.; Derse, D.; Chen, J. Y.; Caradonna, S. J.; Connor, K.; Biochem. Biophys. Acta 1981, 652, 90. (b) McKenna, C. E.; Khawli, L. A.; Bapat, A.; Harutunian, V.; Cheng, Y. C. Biochem. Pharm. 1987, 36, 3103. (c) McKenna, C. E.; Levy, J. N.; Cheng, Y. C.; Starnes, M.; Bapat, A., submitted for publication.

⁽⁶⁾ Hutchinson, D. W.; Naylor, M.; Semple, G. *Chim. Scr.* 1986, 26, 91.
(7) (a) Coleman, J. E. "The Role of Zn(II) in RNA and DNA Polymerases", Chapter 6 In *Zinc Enzymes*; Spiro, T. G., Ed.; Vol. 5 In series *Metal Ions* in *Biology*; Wiley and Sons: New York, 1983; pp 219–252. (b) Wu, F. Y. H.; Wu, C. N. "The Role of Zinc in DNA and RNA Polymerases", Chapter 4 In Zinc and Its Role in Biology and Nutrition; Sigel, H., Ed.; Vol. 5 In the series Metal Ions in Biological Systems; Marcel Dekker, Inc.: New York, 1983; pp 157-192.

⁽⁸⁾ Several non-viral DNA polymerases are now believed not to contain constitutive Zn (ref 9a-f), although metal-binding site(s) are present; on the other hand, Zn has been clearly demonstrated in at least two non-viral RNA polymerases (ref 9g,h).

^{(9) (}a) Walton, K. E.; Fitzgerald, P. C.; Herrmann, M. S.; Behnke, W. D. Biochem. Biophys. Res. Commun. 1982, 108, 1353. (b) Ferrin, L. J.; Mildvan, A. S.; Loeb, L. A. Biochem. Biophys. Res. Commun. 1983, 112, 723. (c) Graham, D. R.; Sigman, D. S. Inorg. Chem. 1984, 23, 4188. (d) Slaby, I.; Lind, B.; Holmgren, A. Biochem. Biophys. Res. Commun. 1984, 122, 1410. (c) Ollis, D. L.; Brick, P.; Hamlin, R.; Xuong, N. G.; Steitz, T. A. Nature (*London*) 1985, 313, 762. (f) Joyce, C. M.; Steitz, T. A. T.I.B.S. 1987, 12, 288. (g) Giedroc, D. P.; Coleman, J. E. Biochemistry 1986, 25, 4969. (h) Mazus, B.; Falchuk, K. H.; Vallee, B. L. Biochemistry 1986, 25, 2941.

^{(10) (}a) Lindson, K.; Deutsch, E.; Barnett, B. L. J. Am. Chem. Soc. 1980, (10) (a) Linkon, R., Denkan, E., Barnet, B. E.J. Mitter, N. Chem. Bor. 102, 2476. (b) Jurisson, S. S.; Benedict, J. J.; Elder, R. C.; Whittle, R.; Deutsch, E. *Inorg. Chem.* **1983**, *22*, 1332. (c) Haromy, T. P.; Knight, W. B.; Donaway-Mariano, D.; Sundaralingam, M. *Inorg. Chem.* **1984**, *23*, 2413.



Figure 3. Structure of [Pt2(cis-dach)2(MDP)]. Distances around the periphery of the Pt are as follows: Pt-N = 2.08 (3), Pt-O = 2.06 (2), O-P = 1.54 (4), Pt-C = 1.83 (2) Å. $[Pt_2(cis-diaminohexane)_2(methy$ lenediphosphonate)]-9H2O crystallizes in the orthorhombic space group blac, with a = 21.67 (1) Å, b = 12.13 (1) Å, c = 23.63 (1) Å, V = 6209(6) Å³, Z = 8. Automated Patterson search techniques were used to obtain the two Pt atoms, and the rest of the structure was refined to final agreement factors of R = 3.6% and R(w) = 3.8% for 1390 reflections.

rophosphate11 complexes, and some preparative12 and solution studies¹³ of related compounds, have been published. In this initial study, we have chosen Pt²⁺ as a useful preliminary model for Zn^{2+} . The well-defined¹⁴ Pt²⁺ square-planar geometry provides a valuable point of departure for evaluation of the less discriminating¹⁴ and more elusive coordination chemistry of Zn²⁺ with phosphonate ligands. This paper describes the preparation and structure determination of three PtII(bisamine)/phosphonate complexes, Na⁺[cis-Pt(NH₃)₂(PFA)]⁻, Na⁺[Pt(dach)(PFA)]⁻, and [Pt2(dach)2(MDP)] (dach = diaminocyclohexane).

Na⁺[cis-Pt(NH₃)₂(PFA)]⁻ was prepared by adding 20 mg of cis-Pt^{II}(NH₃)₂Cl₂ to a 10 mL solution of 20 mg of trisodium phosphonoformate (dark, 50 °C, 12 h), followed by concentration to about 2 mL via rotovaporation. Pale yellow plate-like crystals appeared upon slow evaporation. The structure of the [cis-Pt-(NH₃)₂(PFA)]⁻ anion is shown in Figure 1.¹⁵ The strikingly simple structure of this compound features the expected square-planar geometry of the Pt(II) atom and the phosphoformate ligand forming a five-membered (Pt-O-C-P-O) chelate ring.

Na⁺[Pt(trans-l-dach)(PFA)]⁻ is prepared by adding 20 mg of $Pt(trans-l-dach)Cl_2^{16}$ to 10 mL of an aqueous solution containing 17.9 mg of AgNO₃, followed by 15.8 mg of trisodium phosphonoformate after AgCl was removed. After stirring in the dark for 12 h, the solution (pH = 5.4) was concentrated to 2 mL, and brick-like colorless crystals appeared in about 2 days. The structure of [Pt(dach)(PFA)]⁻ is shown in Figure 2.15 The geometry is very similar to that of the related [cis-Pt(NH₃)₂-(PFA)]⁻ complex, suggesting again a strong tendency for the five-membered (Pt-O-C-P-O) chelate ring to form.

The third complex, [Pt2(dach)2(MDP)], was prepared under similar conditions: 27.0 mg of Pt(cis-dach)Cl2¹⁶ was added to 10 mL of aqueous AgNO₃ (24.1 mg, dark), followed by a 5 mL solution of MDP (12.5 mg; free acid) after AgCl was removed. The pH was adjusted to 7.0 by addition of NaOH, and the mixture was stirred (30 h, room temperature, dark) and concentrated (2.5 mL). Colorless crystals formed in about 10 h upon cooling to 4 °C. The structure of the compound (Figure 3)¹⁵ showed it to be bimetallic, in contrast to the Pt/PFA complexes. This was unanticipated: the preparative procedure described above used a 7547

1:1 ratio of platinum/phosphonate. Each (dach)Pt fragment forms a six-membered (Pt-O-P-C-P-O) ring, and the Pt-Pt distance is nonbonding [3.266 (1) Å].

Our work provides direct confirmation of the proposal⁶ that PFA and MDP form five- and six-membered chelate rings, respectively, with metal (II) ions, at least with Pt(II).¹⁷ Work is currently underway on cognate phosphonate complexes of Zn²⁺, Mn²⁺, and Mg²⁺. Pt^{II} (amine)₂(phosphonate) complexes were also selected as target molecules in recognition of a recent report that the complex cis-[Pt(NH₃)₂(cyclaradine)₂Cl₂]Cl₂ (cyclaradine = carbocyclicarabinofuranosyladenine) and related compounds are active against topical herpes virus infections,18 serving presumably as combination drugs. The possibility that appropriately designed PtL₂(phosphonate)-type complexes might prove to be active against various classes of viruses is being explored.19

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Supplementary Material Available: Tables of atomic coordinates for Na⁺[cis-Pt(NH₃)₂(PFA)]⁻, Na⁺[Pt(dach)(PFA)]⁻, and [Pt₂(dach)₂(MDP)] (2 pages). Ordering information is given on any current masthead page.

(17) It does not, of course, address the issue of which metal ions may be relevant for activity. This question is currently under investigation in our laboratories.

(18) Taylor, R. C.; Ward, S. G.; Schmidt, P. P. U.S. Patent 4 571 335; Chem. Abstr. 1986, 104, 200205y.

(19) Cheng, Y. C.; Bodner, A.; McKenna, C. E.; Bau, R., manuscript in preparation.

Synthesis of the Transient Formyl Complex [CH₃N(PF₂)₂]₃Co₂(CO)(CHO)⁻ and Observation of Facile Formyl Decomposition To Give the Mixed-Valence Complex [CH₃N(PF₂)₂]₃Co₂(CO)₂.

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Metal formyl complexes continue to derive interest as plausible intermediates in the catalytic reduction of carbon monoxide to alcohols and hydrocarbons.1 While many mononuclear formyl complexes have been prepared and isolated,^{2,3} well-characterized polynuclear formyl complexes remain scarce as a result of enhanced decomposition reactivity relative to their mononuclear counterparts.

The major decomposition pathway in polynuclear formyl complexes, of which the dinuclear formyl complex [CH3N(P-

⁽¹¹⁾ Stanko, J. A. In "Platinum Coordination Complexes in Cancer

⁽¹¹⁾ Stanko, J. A. In "Platinum Coordination Complexes in Cancer Chemotherapy", Connors, T. A., Roberts, J. J., Eds.; Vol. 48 In Recent Results in Cancer Research; Springer-Verglag: Berlin, 1974; p.25.
(12) Nepras, M. J., M.S. Thesis, University of South Florida, 1979.
(13) (a) Stunzi, H.; Perrin, D. D. J. Inorg. Biochem. 1979, 10, 523.
(14) Cotton, F. A.; Wilkinson, G. Advanced Inorganic Chemistry; 4th ed.; J. Wiley & Sons: New York, 1980.
(15) The Grand Lamin and Complexe for all these structures described in this.

⁽¹⁵⁾ The final atomic coordinates for all three structures described in this aper are available. See Supplementary Material paragraph at end of paper for ordering details.

^{(16) (}a) Kidani, Y.; Saito, R. Chem. Lett. 1976, 2, 123-126. (b) Kidani, Y.; Inagaki, K.; Iigo, M.; Hoshi, A.; Kuretani, K. J. Med. Chem. 1978, 21, 1315.

^{(1) (}a) Masters, C. Adv. Organomet. Chem. 1979, 17, 61. (b) Henrici-Olive, G.; Olive, S. The Chemistry of the Catalyzed Hydrogenation of Carbon Monoxide; Springer-Verlag: New York, 1984. (c) Blackborow, J. R.; Darodo, R. J.; Wilkinson, G. Coord. Chem. Rev. **1982**, 43, 17. (d) Henrici-Olive, G.;

 ⁽a) Casey, C. P.; Andrews, M. A.; Rinz, J. E. J. Am. Chem. Soc. 1979, 101, 741.
 (b) Casey, C. P.; Andrews, M. A.; Rinz, J. E. J. Am. Chem. Soc. 1979, 101, 741.
 (c) Selover, J. C.; Marsi, M.; Parker, D. W.; Gladysz, J. A. J. Organomet. Chem. 1981, 206, 317.
 (d) Casey, C. P.; Neumann, S. M.; J. Am. Chem. Soc. 1978, 100, 2544.
 (c) Selover, J. C.; Marsi, M.; Parker, D. W.; Gladysz, J. A. J. Organomet. Chem. 1981, 206, 317.
 (d) Casey, C. P.; Neumann, S. M.; Andrews, M. A. Marker, M. M.; Marker, M. M.; Chem. 1980, 206, 206. M. A.; McAlister, D. R. Pure Appl. Chem. 1980, 52, 625. (e) Casey, C. P.;
 Andrews, M. A.; McAlister, D. R.; Jones, W. D.; Harsy, S. G. J. Mol. Catal.
 1981, 13, 43. (f) Casey, C. P.; Neumann, S. M. J. Am. Chem. Soc. 1976, 98, 5395. (g) Casey, C. P.; Andrews, M. A.; McAlister, D. R.; Rinz, J. E. J. Am. Chem. Soc. 1980, 102, 1927.

^{(3) (}a) Lilga, M. A.; Ibers, J. A. Organometallics 1985, 4, 590. Wayland, B. B.; Woods, B. A. J. Chem. Soc., Chem. Commun. 1981, 700. (c) Leoni, P.; Landi, A.; Pasquali, M. J. Organomet. Chem. 1987, 321, 365.
(d) Cohen, H.; Meyerstein, D.; Schustermann, A. J.; Weiss, M. J. Chem. Soc., Chem. Commun. 1985, 424. (e) Smith, G.; Cole-Hamilton, D. J.; Thornton-Pett, M.; Hursthouse, M. B. J. Chem. Soc. Dalton Trans. 1985, 387.